

## ACYLATED FLAVONOL GLYCOSIDES FROM *Sedum aizoon*

Qiu-Yan Luo,<sup>1</sup> Wei-Lin Li,<sup>2\*</sup> and Li-Qiang Wu<sup>2</sup>

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*A new acylated flavonol glycoside (1) and a known compound (2) have been isolated from the whole plant of Sedum aizoon L. Their structures have been established as quercetin 3-O-[β-D-(2'''-O-(E)-feruloyl)-xylopyranosyl]-(1→6)-β-D-glucopyranoside (1) and rutin (2) by means of spectroscopic analysis and chemical methods.*

**Keywords:** *Sedum aizoon*, acylated flavonol glycosides, quercetin 3-O-[β-D-(2'''-O-(E)-feruloyl)-xylopyranosyl]-(1→6)-β-D-glucopyranoside.

*Sedum aizoon* L. is an endemic plant. It is useful for tranquilization, arresting bleeding, and relieving stasis, and as a decoction for counteracting toxicity. The plant has been used to treat pain or trauma and various hemorrhage and palpitation conditions [1, 2]. There are about 150 *Sedum* species distributed in China.

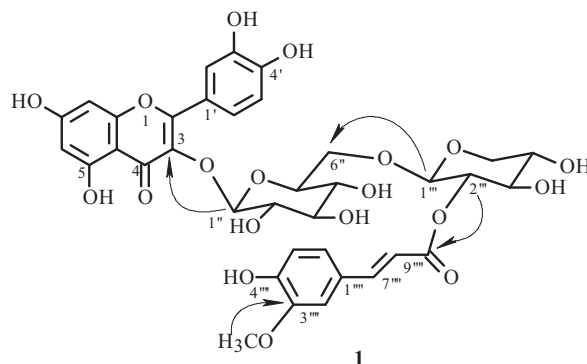
The phytochemical constituents of *Sedum* species, a large genus of the family Crassulaceae, have been extensively reported [3–6]. In our research on this plant, repeated column chromatographic separation of BuOH fractions of the MeOH extract resulted in the isolation of a new flavonol glycoside (**1**), as well as a known flavonol glycoside (**2**), whose structure was determined to be rutin (**2**) [7] by comparing its spectroscopic data with those in the literature.

Compound **1** (see structure in Fig. 1) was isolated as a yellowish amorphous powder. The HR-ESI-MS showed a quasi-molecular ion peak at  $m/z$  773.1929  $[M + H]^+$  (calcd for 773.1937), corresponding to the molecular formula of  $C_{36}H_{36}O_{19}$ . The IR spectrum indicated the presence of hydroxyl groups, an ester carbonyl group, and an aromatic ring. Positive ESI-MS/MS of **1** showed a quasi-molecular ion  $[M + H]^+$  at  $m/z$  773.2 and fragment ions at  $m/z$  641.2  $[M + H - 132]^+$  and 479.2  $[M + H - 132 - 162]^+$ , indicating the loss of pentosyl and hexosyl moieties from the quasi-molecular ion. The molecular weight of the aglycone was  $m/z$  302. Its  $^1H$  NMR spectrum showed the presence of one chelated hydroxyl group (13.04, s, 5-OH), three ABX protons belonging to a 3,4-disubstituted phenyl group ( $\delta$  7.60, 1H, d,  $J = 1.8$  Hz, H-2'; 6.89, 1H, d,  $J = 8.0$  Hz, H-5'; 7.56, 1H, dd,  $J = 1.8, 8.0$  Hz, H-6'), two protons typical of a 5,7-disubstituted ring A ( $\delta$  6.45, 6.84, each 1H, d,  $J = 2.0$ , H-8 and H-6), and two anomeric protons representing two glycosyl units ( $\delta$  5.15, d,  $J = 7.5$  Hz and 4.43, d,  $J = 7.5$ ). These data, together with those of  $^{13}C$  NMR spectroscopy, indicated the presence of a quercetin [8] and two glycosyl units [9]. After acid hydrolysis, quercetin was obtained as the aglycone. Glucose and xylose moieties were determined via acid hydrolysis and comparing with standard D-glucose and D-xylose. From the  $^1H$  NMR spectrum, the coupling constant of Glc H-1'' ( $J = 7.5$  Hz) and Xyl H-1''' ( $J = 7.5$  Hz) suggested a  $\beta$ -anomeric configuration for the glucose and xylose. The cross-peak between C-3 and H-1'' in the HMBC spectrum established the linkage point at quercetin and the glucosyl moiety. From the  $^{13}C$  NMR spectral data, the C-6'' (Glc) was shifted downfield at  $\delta$  68.9, indicating that glycosylation was at this unit. In the HMBC spectrum, the cross-peak between C-6'' (Glc) and H-1''' (Xyl) established the linkage point between the two sugar moieties (Fig. 1). The presence of the feruloyl moiety was deduced from the ABX spin system at  $\delta$  6.63 (1H, d,  $J = 8.0$  Hz, H-5'''), 6.91 (1H, dd,  $J = 1.8, 8.0$  Hz, H-6'''), and 7.14 (1H, d,  $J = 1.8$  Hz, H-2'''). The *E*-configuration of the double bond was confirmed by the large coupling constant of the coupled olefinic protons at  $\delta$  7.33 (1H, d,  $J = 15.9$  Hz, H-7''') and 6.27 (1H, d,  $J = 15.9$  Hz, H-8'''). The methoxy group at  $\delta$  3.85 (3H, s,  $OCH_3$ ) was determined at C-3'''' of the feruloyl, which was evidenced by the cross-peaks of the *O*-methyl hydrogens to C-3'''' ( $\delta$  147.9) in the HMBC spectrum (Fig. 1).

1) Department of Gynecology and Obstetrics, the Third Affiliated Hospital to Xinxiang Medical University, Xinxiang 453003, P. R. China; 2) Department of Medicinal Chemistry, Pharmacy College of Xinxiang Medical University, Xinxiang 453003, P. R. China, fax: +86 373 302 98 79, e-mail: liweilin2002@163.com. Published in *Khimiya Prirodnykh Soedinenii*, No. 1, pp. 26–28, January–February, 2012. Original article submitted December 25, 2010.

TABLE 1.  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Chemical Shifts of **1** (DMSO- $d_6$ ,  $\delta$ , ppm, J/Hz)

C atom	$\delta_{\text{H}}$	$\delta_{\text{C}}$	C atom	$\delta_{\text{H}}$	$\delta_{\text{C}}$
2		157.8	Xyl		
3		136.4	1'''	4.43 (d, J = 7.5)	104.7
4		179.5	2'''	4.62 (dd, J = 9.0, 9.0)	73.5
5		162.5	3'''	3.30 m	74.6
6	6.45 (d, J = 2.0)	99.8	4'''	3.40 m	69.8
7		163.1	5'''a	3.76 (dd, J = 12.0, 2.0)	65.8
8	6.84 (d, J = 2.0)	94.5	5'''b	3.04 (dd, J = 12.0, 4.0)	
9		157.6	<i>trans</i> -Feruloyl		
10		107.9	1''''		125.5
1'		122.6	2''''	7.14 (d, J = 1.8)	110.7
2'	7.60 (d, J = 1.8)	117.4	3''''		147.9
3'		145.8	4''''		145.3
4'		149.2	5''''	6.63 (d, J = 8.0)	115.3
5'	6.89 (d, J = 8.0)	115.1	6''''	6.91 (dd, J = 8.0, 1.8)	123.4
6'	7.56 (dd, J = 8.0, 1.8)	123.0	7''''	7.33 (d, J = 15.9)	144.6
3-O-Glc		102.7	8''''	6.27 (d, J = 15.9)	115.3
1''	5.15 (d, J = 7.5)	73.7	9''''		168.7
2''	3.34 m	77.4	OMe	3.85 s	56.2
3''	3.30 m	70.3			
4''	3.10 m	76.2			
5''	3.58 m	68.9			
6''a	3.96 (dd, J = 12.2, 3.0)				
6''b	3.62 (dd, J = 12.2, 5.0)				

Fig. 1. Key HMBC correlations of compound **1**.

The connection of the xylopyranosyl group through C-2''' to the carbonyl group of *trans*-feruloyl at C-9'''' was determined by the HMBC correlations, where the cross-peaks of H-2''' ( $\delta$  4.62) to C-9'''' ( $\delta$  168.7) were presented. Thus, the structure of **1** was elucidated to be quercetin 3-O- $[\beta$ -D-(2''''-O-(*E*)-feruloyl)-xylopyranosyl]-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside.

## EXPERIMENTAL

**General Experimental Procedures.** Melting points were determined on an XT-4 digital melting-point apparatus with a binocular microscope and are uncorrected. UV and IR spectra were recorded respectively with a Jasco V-650 spectrophotometer and an FTS-40 infrared spectrometer with KBr pellets. HR-ESI-MS was measured on an Agilent 6520 Accurate-Mass Q-TOF LC/MS system.  $^1\text{H}$  NMR (400 MHz) spectra were recorded on Bruker-400 spectrometer.  $^{13}\text{C}$  NMR (100 MHz) and HMBC spectra were run on a Bruker-400 spectrometer with TMS as internal standard. Silica gel (Qingdao Marine Chemical Factory, 200–300 mesh) and Sephadex LH-20 (Pharmacia) and RP-18 (Merck, 40–60  $\mu\text{m}$ ) were used for column chromatography (CC). Reversed-phase HPLC separations were performed on a Shimadzu LC-6A system, detected by a UV detector at 254 nm and equipped with a YMC semipreparative  $\text{C}_{18}$  column (10  $\mu\text{m}$ , 10  $\times$  250 mm) running with a flow

rate of 4 mL/min. Precoated plates of silica gel GF<sub>254</sub> and silica gel RP-18 F<sub>254</sub> (Merck) were used for TLC, and spots were detected under UV light or by heating after spraying with 98% H<sub>2</sub>SO<sub>4</sub>-EtOH (10:90, v/v).

**Plant Material.** The plant *S. aizoon* was collected from Taihang Mountain, Henan province, P. R. China, in September 2007 and identified by Prof. Chen-Ming Dong, Henan College of Traditional Chinese Medicine. A voucher specimen (No. 200798) had been deposited at the Herbarium of the Pharmacy College of Xinxiang Medical University.

**Isolation.** The dry whole plant tissue of *Sedum aizoon* L. (4.5 kg) was macerated at room temperature with MeOH for 48 h four times (4 × 10 L, 7 day) and filtered. The combined methanolic extract was concentrated *in vacuo* to yield a crude extract (350 g), which was dissolved in water (2.0 L) and then partitioned with petroleum ether, EtOAc, and *n*-BuOH successively. The petroleum ether layer was evaporated to give 25.4 g residue; the EtOAc fraction yielded 95.4 g, and the *n*-BuOH fraction yielded 230 g. The *n*-BuOH fraction was passed over a D<sub>101</sub> macroporous resin CC (1 kg) eluted with a gradient of aqueous EtOH (0, 30, 60, 80, 95, v/v) to yield five fractions (1–5). Fraction 3 (11.3 g) was subjected to reversed-phase silica gel CC and eluted with a gradient of aqueous MeOH (0, 30, 45, 60, 75, 95, v/v) to give six fractions (3.1–3.6), Fraction 3.2 (890 mg) was subjected to Sephadex LH-20 CC (eluted with MeOH) and preparative HPLC (60% aqueous MeOH, 4 mL/min) to yield compound **1** (7.8 mg); fraction 3.3 (1.35 g) was also purified by Sephadex LH-20 CC (eluted with MeOH) to give compound **2** (10 mg).

**Compound 1.** Yellowish amorphous powder (MeOH), mp 201.3–202.1°C.  $[\alpha]_{\text{D}}^{20}$  –62° (*c* 0.2, MeOH). UV (MeOH,  $\lambda_{\text{max}}$ , nm): 256, 268, 328. IR (KBr, v, cm<sup>-1</sup>): 3434 (br), 2918, 1696, 1654, 1605, 1510, 1445, 1359, 1263, 1067, 812, and 595. For <sup>1</sup>H NMR and <sup>13</sup>C NMR data, see Table 1. ESI-MS *m/z*: 773.19 [M + H]<sup>+</sup>, 795.18 [M + Na]<sup>+</sup>; HR-ESI-MS *m/z*: 773.1929 [M + H]<sup>+</sup> (calcd for C<sub>36</sub>H<sub>36</sub>O<sub>19</sub>, 773.1937).

**Acid Hydrolysis of Compound 1.** A solution of compound **1** (2.0 mg) in 1 M HCl (1.0 mL) was heated under reflux for 3 h. After cooling, the reaction mixture was extracted with EtOAc, and the residues from the organic phase were identified as quercetin and ferulic acid by comparative TLC [Si gel, CHCl<sub>3</sub>-MeOH (8:2)] with authentic samples available in our laboratory. The aqueous phase was concentrated, and the sugar was identified as D-glucose and D-xylose by TLC [Si gel, BuOH-AcOH-H<sub>2</sub>O (3:1:1)] with a standard sample (Merck).

**Compound 2.** Yellow amorphous powder, mp 185–188°C. ESI-MS (*m/z*): 611 [M + H]<sup>+</sup>. <sup>1</sup>H NMR (400 MHz, acetone-d<sub>6</sub>,  $\delta$ , ppm, J/Hz): 12.58 (1H, s, 5-OH), 7.54 (1H, d, J = 1.8, H-2'), 7.52 (1H, dd, J = 8.4, 1.8, H-6'), 6.84 (1H, d, J = 8.4, H-5'), 6.37 (1H, d, J = 1.5, H-8), 6.18 (1H, d, J = 1.5, H-6), 5.34 (1H, d, J = 7.2, H-1''), 4.37 (1H, d, J = 1.6, H-1'''), 0.98 (3H, d, J = 6.0, H-6''').

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